

DOI 10.24425/pjvs.2023.145033

Review paper

Alternatives to zinc oxide in pig production

Z. Pejsak¹, P. Kaźmierczak², A.F. Butkiewicz², J. Wojciechowski³,
G. Woźniakowski⁴

¹University Center of Veterinary Medicine JU-AU, Mickiewicza Avenue 24/28, 30-059 Krakow, Poland

²Institute of Veterinary Medicine,

Nicolaus Copernicus University in Toruń, Lwowska 1, 87-100 Toruń, Poland

³Private Veterinary Practice, Grabowa 3, 86-300 Grudziadz, Poland

⁴Department of Infectious and Invasive Diseases and Veterinary Administration,
Institute of Veterinary Medicine, Nicolaus Copernicus University in Toruń, Lwowska 1, 87-100 Toruń, Poland

Abstract

Zinc oxide (ZnO) has been applied for many years in the production of pigs to reduce the number of diarrhoea in weaned piglets. In June 2022, the European Union banned the use of zinc oxide (ZnO) in pig feed. According to scientific reports, the main reason was the accumulation of this microelement in the environment of pig production. It has been shown that frequent application of ZnO can lead to increased antibiotic resistance in pathogenic swine microflora. The main alternatives to ZnO are probiotics, prebiotics, organic acids, essential oils, and liquid feeding systems.

Alternatives to ZnO can be successfully used in pig production to reduce the number of diarrhoea among piglets during the postweaning period. Additional reports indicated that bacteriophage supplementation has a positive effect on the health of pigs. The article provides an overview of current ZnO substitutes that can be used in pig farming.

Keywords: zinc oxide, alternatives, probiotics, organic acids, liquid feeding, bacteriophages

Introduction

Zinc represents a key element for the functioning of all living beings, as well as for the proper course of biological processes at the cellular level (Broadley et al. 2007, Prakash et al. 2015). This element is involved in actions of several enzymes and transcription factors (TFs) (Broadley et al. 2007). Zinc and especially zinc oxide (ZnO) have played an important role in pig production until the European Union (UE) has introduced

a limitation on the use of antibiotics as feed additives, including natural growth promoters (AGP) and metaphylaxis (2006) (Casewell et al. 2002). According to the scientific reports and opinions of leading food safety authorities, ZnO has an adverse effect on the environment due to its accumulation and potential increase in resistance of certain bacterial species (European Commission 2003, Bednorz et al. 2013, Vahjen et al. 2015, European Commission 2017).

The exact way in which ZnO limits porcine diarrhea

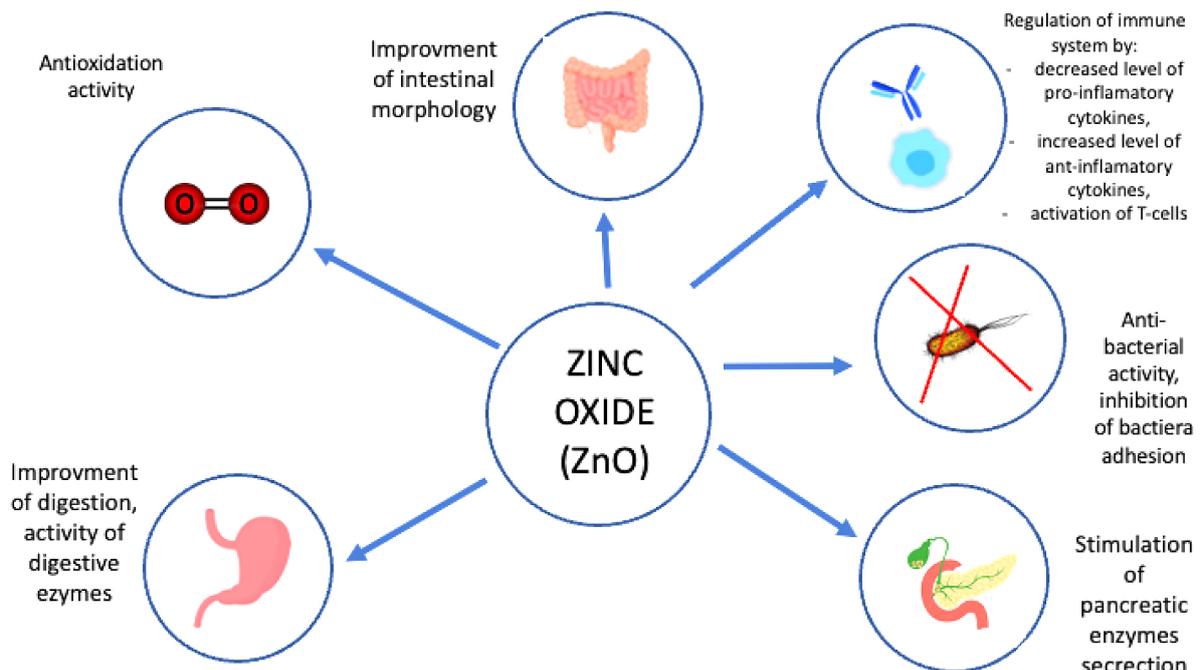


Fig. 1. Influence of zinc oxide on the functions of the digestive system and the activity of the immune system.

is not yet fully understood. Although previous studies have assumed that ZnO activity is related to the factors presented in Fig. 1. It seems that ZnO's primary mechanism of action is related to improve nutrient absorption and intestinal morphology, rather than simply fulfilling daily nutritional requirements (Pearce et al. 2015). Additional research has confirmed that pharmacological supplementation with ZnO improves performance by increasing the villi: crypt ratio, including protein levels, and promoting enterocyte proliferation (Grilli et al. 2015, Zhu et al. 2017). ZnO also enhances the expression of tight junctions, which reduces intestinal permeability, a crucial factor during a delicate phase for piglets that can lead to diarrhea (Zhang and Guo 2009). The antioxidant properties of ZnO may also play a role in its positive effect on the intestinal mucosa (Zhu et al. 2017).

Although ZnO has been shown to cause oxidative stress in microbial cells (Pasquet et al. 2014, Lee et al. 2018) its direct antimicrobial activity against *Escherichia coli* F4, the primary cause of porcine diarrhoea, is limited (Söderberg et al. 1990). Moreover, ZnO supplementation is mainly directed against Gram-positive bacteria rather than Gram-negative bacteria, with ZnO-supplemented animals showing higher coliforms and enterococci, but lower lactic acid bacteria (Söderberg et al. 1990, Højberg et al. 2005). Therefore, mechanisms other than direct antimicrobial activity may exist for ZnO.

ZnO has been shown to protect cultured enterocytes from ETEC-induced damage and inhibit bacterial

adhesion to cells (Roselli et al. 2003). It also modulates cytokine expression, improving the transcription of the anti-inflammatory cytokine TGF- β while reducing the expression of several pro-inflammatory cytokines (Sargeant et al. 2011, Grilli et al. 2015, Zhu et al. 2017, Gammoh and Rink 2019). ZnO has a key action on the immune system (Gammoh and Rink 2019), promoting the development and function of macrophages (Ercan and Bor 1991), natural-killer lymphocytes (Rolles et al. 2018), and T helper cells (Honscheid et al. 2009). In weaned piglets, pharmacological doses of ZnO increase the number of regulatory T cells, modulating the immune response and maintaining homeostasis (Kloubert et al. 2018). ZnO can also stimulate ghrelin secretion and increase the activity of pancreatic digestive enzymes, leading to higher nutrient digestibility and improved systematic performance in weaning piglets (Yin et al. 2009, Mac Donald 2000, Hedemann et al. 2006).

ZnO partially solved problems in pig production associated with diarrhea in weaned piglets. Weaning is currently carried out in most farms between 21 and 28 days after farrowing (Jensen and Recén 1989, Bøe 1991). The current knowledge indicates the positive effects of early weaning of piglets from the age of 3 weeks. During the weaning process, there are several morphological, enzymatic, and physical changes in piglet organism, which increase the risk of gastrointestinal tract disorders, especially diarrhea. Piglet diarrhea leads to decrease in economically important production rates in farms (Rhouma et al. 2017). The use

of ZnO after weaning has been shown to improve intestinal flora (Hu et al. 2013, Hu et al. 2015) and significantly reduces mortality, relieves symptoms, and improves breeding parameters such as weight gain and feed intake (Poulsen 1998, Case and Carlson 2002).

Zinc is a one of the heavy metals with low bioavailability (European Commission 2003) and must be administered in high doses (2000-4000 ppm) to allow optimal absorption in the small intestine of piglets. As a result of poor absorption, most zinc is excreted through urine and feces, leading to accumulation in the environment, similarly to other heavy metals such as mercury or lead (Ociepa-Kubicka and Ociepa 2012).

There are numerous reports regarding antimicrobial resistance caused partially by ZnO administration. For instance, the study conducted by Bendorz et al. (2013) showed a probable role of ZnO in *Escherichia coli* antibiotic resistance. Similar observations were made in case of *Staphylococcus aureus*, where the possibility of the increased resistance to methicillin caused by ZnO administration was demonstrated. Vahjen et al. (2015) concluded that high supplementation with ZnO in piglet diet may increase the appearance of resistance genes among Gram-negative bacteria to tetracyclines and sulfonamides.

The aim of this paper is to provide the current review on pig supplements and alternatives that may replace the effect of ZnO administration in feed for piglets.

Low protein concentration in feed

The post-weaning time represents one of the most stressful times for piglets. One of the most important stressing factors is represented by a change from colostrum to solid feed. The potential problems are associated with low feed intake, non-effective digestion, lack of micro- and microelements, anorexia as well as immune deficiency. Additionally in weaned piglets, the HCl production is too low to ensure the proper digestion of regular feed (Halas et al 2007). In general, the protein concentration in feed for this age group exceeds 200 g/kg of feed, which may overpass the capacity of piglet stomach and proper digestion. The undigested protein that concentrates in the large intestine leads to diarrhea and replication of pathogenic microflora including enterotoxigenic *E.coli* (ETEC) (Nyachoti et al. 2006, Wellock et al. 2008, Pieper et al. 2016). On the other hand, the accumulation of toxic fermentation metabolites, excessive growth of pathogenic microflora as well as not fully developed immune system of piglets may lead to diarrhea and increased mortality. Therefore, it is very important to carefully

choose the feed considering the protein content, including individual amino acids, and to plan the moment of introduction of solid feed for unweaned piglets (Yue and Qiao 2008, Byrgesen et al. 2021).

It has been shown that piglets fed containing from 15 to 18% of protein showed better resistance to *E. coli* F4 infection during the animal trial (Yue and Qiao 2008, Heo et al. 2010). A low-protein diet can reduce diarrhea symptoms up to 30% in piglets up to 15 kg of body weight. Furthermore, some researchers pointed out that the levels of IL-1, TNF- α , IL-2, IL-6, and IL-10 and IL-13 in the colonic mucosa of weaned piglets decreased, which is most likely due to lower concentration of protein in the feed (Wan et al. 2020).

However, low weight gain and amino acid deficiency can be a serious problem in a low protein diet in piglets (Yue and Qiao 2008). Several studies have shown that threonine, arginine, glutamine, methionine, and cysteine are responsible for the proper functioning of the intestinal immune barrier, anti-inflammatory and antioxidant functions, and stress reduction after weaning. Methionine, lysine, threonine, and glutamate also influenced the normal growth and composition of the intestinal bacterial flora (Wang et al. 2009, Liao 2021, López-Gálvez et al. 2021). Threonine is an essential component of mucins contained in intestinal mucin, since it represents between 28% and 35% of amino acids in mucinous proteins. Stimulation of the immune system, e.g., in case of *E. coli* infection, increases mucus production as well as requirement for threonine in piglets (Lien et al. 1997, Le Floc'h and Seve 2007, Trevisi et al. 2015, McGilvray 2019). The results of other studies indicate that a 2 g/kg of threonine feed supply as recommended by the National Research Council (NRC 2012) improves the intestinal barrier function and intestinal morphology, increasing the probability that piglets avoid diarrhea (Ren et al. 2014, Zhang et al. 2019, Koo et al. 2020, Table 1).

Currently, various concepts are being used to create a scheme for the "ideal protein." One of the more well known is InraPorc which acts as a tool for determining the ratio of amino acids relative to the standardised ileal digestible (SID) lysine (Lys=100%) (Van Milgen et al. 2008, Van Milgen and Dourmad 2015, Dourmad et al. 2008).

Therefore, when choosing a low-protein diet, supplementation with amino acids is recommended to avoid deficiencies that affect animal performance (Halas et al. 2007).

Probiotics

Probiotics are living microorganisms that reach the intestine in sufficient quantity and active to induce

Table 1. Expressed as a percentage of the standardised ileal digestible (SID) Lys requirement (Lys=100%) (Dourmad et al. 2008, Van Milgen et al. 2008, Van Milgen and Dourmad 2015).

Items	Growing pigs 20-140 kg	Gestating sows	Lactating sows
Lys (base)	100	100	100
Met	30	28	30
Met+Cys	60	65	60
Thr	65	72	66
Trp	18	20	19
Val	70	75	85
Ile	55	65	60
Leu	100	100	115
Phe	50	60	60
Phe+Tyr	95	100	115
His	32	30	42
Arg	42	-	-

a positive effect on health of the host, and improving the functioning of the digestive system. The microorganisms contained in probiotics should not contain pathogenic, toxic, or antibiotic resistant strains. The colonization of the intestine by microorganisms contained in probiotics has a few advantages including maintaining the integrity of the intestinal barrier, competitiveness of the pathogenic microflora, production of antimicrobial compounds and inactivation of toxins. Other indirect advantages include stimulation of host's immune response and improvement of performance, better weight gain, reduction of animal stress, and reduction of methane emissions (Dianawati 2016, Dubreuil 2017, Lan 2017, Wang 2017, Barba-Vidal et al. 2019). Currently there are number probiotics for pigs available on the market, containing *Bacillus* spp., *Bifidobacterium* spp., *Clostridium* spp., *Enterococcus* spp., *Lactobacillus* spp. and *Pediococcus* spp. Parallely yeast probiotics belonging to the species *Saccharomyces cerevisiae* are widely used. Commercial products generally contain different strains of one or more types of bacteria and/or yeast. They are applied as feed additives in quantities between 10^8 and 10^{11} colony formation units per kg of feed (CFU/kg) (López-Gálvez et al. 2021).

Studies have shown that *Bacillus* spp. preparations have a positive effect on intestinal flora by the reduction of diarrhea incidence and improved body growth of weaned piglets. Supplementation with *B. subtilis* (10^9 CFU/kg of feed) reduced diarrhea caused by the enterotoxigenic *E. coli* (ETEC) F18 fimbriae. From a clinical point of view, *B. subtilis* and *B. licheniformis* are the most frequently used probiotics. It has been shown that the supplementation with *B. subtilis* KN-42 at a dose of 2.0×10^9 and 4.0×10^9 CFU/kg feed had a comparable

effect that caused by application of neomycin, especially in piglets during the first 14 days post-weaning (Hu et al. 2014). Similarly, *B. licheniformis* ATCC 21424 at dose of 1.6×10^9 CFU spores/g of feed or 4.8×10^9 CFU spores/g of feed reduced the prevalence of piglet diarrhea also (Dumitru et al. 2020). Both species stimulated the growth of *Lactobacillus* spp. and reduced the amount of *E. coli* in the intestines. The combination of several strains of *Bacillus* spp. during the studies did not provide the same results as their separated administration (Zhang et al. 2017).

Bifidobacterium spp. bacteria are rods that physiologically colonize the intestinal flora in pigs. They are responsible for fermentation of glucose into lactic acid, which lowers the pH value. The study conducted by Rhouma et al. (2017) showed that *B. lactis* HN019 supplementation (10^9 CFU/Ferkel) may inhibit rotavirus A (RVA) and *E. coli* caused diarrhea. Supplementation with a multi-strain probiotic *B. longum* subsp. *infantis* CECT 7210 or *B. animalis* subsp. *lactis* BPL6 (1.0×10^9 CFU) had a positive effect on the reduction of infection with *Salmonella typhimurium* in piglets, what was observed as a decreasing incidence of diarrhea and increasing feed intake (Barba-Vidal 2017).

Enterococci also have a positive effect on intestinal functions in weaned piglets. The conducted experiments pointed out that administration of *E. faecium* R1 (6.5×10^6 CFU/g feed) had a positive influence onto the reduction on diarrhea symptoms in piglets. The bacteria stimulated renewal of the intestinal mucosa, appetite, and detoxication (Zeyner and Boldt 2006, Hu et al. 2015, Zhang et al. 2021).

Administration of *Lactobacillus plantarum* ZJ316 (1×10^9 CFU/day), *L. plantarum* PFM105 (2×10^7 CFU/g feed) or a combination of *L. plantarum* and *L. reuteri*

(after 2×10^8 CFU/g feed) via drinking water resulted in a similar effect that caused by antibiotic administration (Suo et al. 2012, Wang et al. 2019).

Many researchers believe that supplementation with a nonpathogenic strain of *E. coli* carries a relatively high risk of diarrhea. However, it has been shown that *E. coli* administration is effective to reduce inflammation of intestines in pigs. This is because the presence of the pathogenic and non-pathogenic strains of *E. coli* within the common biological niche (Canibe et al. 2022). Furthermore, the study conducted by Hrala et al. (2021) showed 100% effectivity in diarrhea inhibition during administration of three human *E. coli* strains including 582, B771 and B1172 at the dose of 1.0×10^9 CFU. However, there still little is known about the positive effect of *E. coli* on the pig organism.

As it has been mentioned, yeasts, in particular *S. boulardii* mafic-1701 at the dose of 1.0×10^8 CFU/kg feed), *S. cerevisiae* (2.0×10^8 CFU/ml, 10 ml/day), *Candida utilis* (1.0×10^9 CFU/ml, 1 ml per day), improve growth parameters in weaned piglets and reduce diarrhea. The applied mixture of probiotics reduced mortality of piglets, increased the diversity of intestinal flora, and had an antioxidant effect. Supplementation of *C. utilis* may be enhanced with *Yucca schidigera* extract. This combination has been shown to have a better effect than administration *Candida utilis* alone (Zhang et al. 2020, Zhaxi et al. 2020, Yang et al. 2021).

In conclusion, application of probiotics has several benefits and should be considered as an alternative to zinc oxide and antibiotics. However, administration of probiotics in excessive doses may significantly alter intestinal flora and affect the immune system in pigs (Canibe et al. 2022).

Organic acids

Application of organic acids in pig farming has been found to lower the pH of the digestive tract and reduce the incidence of diarrhea in weaned piglets, especially during the change from liquid to solid feed (Suiryanrayna and Ramana 2015). Short-chain acids such as formic acid, acetate acid, propionic acid, lactic acid, malic acid, oxalic acid, citric acid, as well as fumaric and sorbic acid are the best alternatives to antibiotics or zinc oxide. The additive of organic acids to feed introduces a sour taste, which is attractive to pigs (Woźniakowska et al. 2017). Organic acids have antibacterial and antifungal properties and are applied by many farmers to acidify drinking water. The addition of 0.5% of citric acid to feed for piglets reduced the infection with *Salmonella typhimurium* causing inflam-

mation of small intestines and colon caused by the *E. coli* serotype KCTC 2571 (Ahmed et al. 2014). Application of organic acids leads to an increase in proteolytic enzymes activity enabling better protein digestion. Acidification leads to the transformation of the pepsinogen proenzyme into endopeptidase, a pepsin that is necessary for proper functioning of the digestive system. Organic acids may influence the morphology of mucosa and stimulation of the pancreatic secretion and serve as substrates in metabolic pathways. The frequent disease in weaned piglets is oedema disease (*Morbus oedematosus*) caused by the *E. coli* serotypes K88 (F4) or F18, which produce Stx2e toxin. The poison damages walls of small blood vessels, including those in the brain, and causes fluid retention or swelling. Vascular damage is associated with characteristic neurological symptoms (Moxley 2000). The studies conducted by Tsilotiannis et al. (2001) showed that application of citric acid in piglets leads to significantly lower mortality. They also pointed out that application of organic acids may be useful in control or prevention of odema disease. In summary, the use of organic acids in pig feed allows the reduction of intestinal bacteria, especially *E. coli*, stimulates the immune system, improves digestibility parameters, and reduces the occurrence of diarrhea in piglets. Therefore, they present very strong alternative zinc oxide (Bonetti et al. 2021).

Polyphenols from plant extracts

Polyphenols are among the biologically active substances that are present in large quantities in vegetables and fruits. Polyphenols include flavonoids, tannic acid, and ellagitannin (Quideau et al. 2011). Polyphenols used as food supplements are generally assumed to improve the function of the digestive system and are involved in metabolic pathways associated with the intestinal flora (Del Rio et al. 2013, Catalkaya et al. 2020). It is believed that an economically and health-optimal source of polyphenol compounds may be fruit pomace, which are by-products of the food industry, which could be frequently found in animal farms. The results of recent studies showed that the dried grape marc as a 5% of the feed had a positive effect on young piglets (Catalkaya et al. 2020). Both the polyphenols present in the pomace and their metabolites can be detected in the intestines of pigs fed with the enriched feed. The action mechanism of phenols affects the cattle digestive tract similarly to growth promoters increasing the secretion of digestive enzymes and reducing several unwanted bacteria in the gastrointestinal tract or modulating intestinal morphology due to their antioxidants

and anti-inflammatory effects. Similar to organic acids, polyphenols in sufficient concentration promote pigs interest in feed due to their attractive taste (Mahfuz et al. 2021). Luna et al. (2010) investigated the effects of polyphenols on the quality of poultry meat. Thymol and carvacrol are polyphenols that occur naturally in thyme (Bouchra et al. 2003, Goodner et al. 2006). The authors pointed out that these additives can be used to improve meat quality (Luna et al. 2010). It has been also reported that phenol compounds may increase fat metabolism. Supplementation with phenol compounds has been believed to stimulate sterol excretion in feces and reduce lipid absorption resulting in lower plasma and liver cholesterol levels (Park et al. 2002). Chrisaki et al. (2012) have reported that dietary supplementation with phenol carriers can effectively inhibits *E.coli* and *Clostridium perfringens* colonization due to the antimicrobial properties of existing phenol compounds. Considering these features of polyphenols they may present an alternative to zinc oxide or antibiotics in the future. More studies on these substances in pig feed are needed to clearly determine whether a food supplement containing polyphenols could replace tried and tested remedies and which ones.

One of the current scientific project conducted by one of the authors of this paper (prof. Grzegorz Woźniakowski) is focused on examination of natural plant extracts influence on replication inhibition of an economically important coronaviruses infecting pigs. As it turns out, an effective solution is offered by a modern method of supercritical extraction with carbon dioxide in a supercritical state (scCO₂, supercritical fluid extraction – SFE), which guarantees the extraction of biologically active compounds without losing their properties and is completely neutral to the environment, plants and animals. Supercritical fluid extracts of plant raw materials are mixtures of several active substances including polyphenols. As the SFE is considered appropriate for the extraction of lipid compounds mainly, the addition of polar co-solvents (water, ethanol) to carbon dioxide widens the possibility of the extraction of polar substances (polyphenols).

The multiple study on natural extracts obtained from fruits, herbs, algae and lichens against human and animal viruses showed the inhibiting potential of the extracts on the replication of the viruses *in vitro* and *in vivo*. Taking into account the antiviral and biocidal potential of natural plant extracts as well as the technological progress in chemical sciences including supercritical extraction with carbon dioxide the efficient procedures have been developed to obtain antiviral compounds including phenols, terpenes, flavonoids and flavones from numerous plants. The antiviral extracts

have been obtained from chokeberries (*Aronia melanocarpa*), cloves (*Syzygium aromaticum*), raspberries (*Rubus idaeus*) as hop (*Humulus lupulus*). The main goal of the currently ongoing studies is to determine to single and synergistic effect of 4 natural extracts against porcine epidemic coronavirus PEDV. Meanwhile, in countries with intensive pig production, including the United States of America or the Republic People of China, losses caused by the occurrence of PEDV reached hundreds millions of US dollars. Due to the very high contagiousness and morbidity of porcine coronaviruses, especially PEDV the most important is the prevention of swine infection by the effective biosecurity rules implementation. Therefore, the influence of natural extracts on PEDV replication inhibition *in vitro* and *in vivo* is going to be performed within the realized project.

Essential oils

Essential oils are extracts of plant origin that are produced as secondary metabolites stored by the plant in secretion cells, cavities, ducts, or epidermis cells. They usually have a typical smell or taste of the plant from which they originate (Nazzaro et al. 2013, Puvača et al. 2013). The biological activity of oils depends on the number of active substances and the relationship between them. For many years, essential oils have been attributed an antimicrobial effect due to a cascade of reactions that affect the entire bacterial cell. In addition, they not only inhibit the growth of bacteria or fungi, but also inhibit the production of their toxic metabolites (Burt and Reinders 2003, Chorianopoulos et al. 2008, De Martino et al. 2009).

In pig farming, essential oils can be used as antibacterial, antioxidant, anti-inflammatory, and anticoccidial agents. They improve the palatability, odor, and digestibility of food (Kroismayr et al. 2006, Brenes and Roura 2010, Sutaphanit and Chitprasert 2014). The conducted studies have shown that cinnamon, thyme, and clove oil are among the most effective extracts against Gram-negative and Gram-positive bacteria, fungi, and yeasts (Abbaszadeh et al. 2014). The MICs of the cinnamon oil and thyme oil components against *E. coli* and *S. typhimurium* DT104 ranged from 100 to 140 µg/ml or µl/ml. In comparison, MICs of clove oil between ranged between 230 and 300 µg/ml or µl/ml (Si et al. 2006). Since the MIC values of the oils are comparable to those of the antibiotics used in pig production, essential oils may be a very good substitute to them. In addition, *Lactobacillus plantarum* and *L. acidophilus* are significantly less sensitive to plant extracts (Zeghib et al. 2017). Other studies suggested

that essential oil supplementation may increase the number of *Lactobacillus* spp. and its biodiversity (Zeng et al. 2015, Li et al. 2017, Wei et al. 2017).

Piglets and weaners are very susceptible to stressors such as weaning, heat stress, overcrowding or transport. This results in a cascade of reactions leading to oxidative stress, which in turn leads to a decrease in the performance of the animals, a decrease in immunity, appetite, the occurrence of diarrhea, and miscarriages in sows. Many studies pointed to the antioxidant effect of essential oils (Tan et al. 2015, Zou et al. 2016, Baschieri et al. 2017, Liu et al. 2017). Zou et al. (2016) have shown that oregano oil in the porcine intestine significantly reduces reactive oxygen and malondialdehyde, which are responsible for oxidative stress (Zou et al. 2016). The same essential oil that was administered to pregnant sows as well as during lactation period improved the performance of piglets by reduction of the oxidative stress in the sows (Tan et al. 2015).

It is possible to use different oils at the same time without antagonistic effect between them. The addition of cinnamon and thyme oil ingredients (100 mg/kg) for piglets from birth to the age of 28 days leads to tangible results. After 14 days of supplementation, the average daily body mass increase, food intake, and the incidence of diarrhea, were comparable to those caused by antibiotics or ZnO. In addition, intestinal morphology improves and oxidative stress decreases (Li et al. 2012, Tian and Piao 2019).

Liquid feeding

Two different feeding systems are distinguished in pig production. The first one is dry feeding, while the second is liquid feeding (Byrgesen et al. 2021). The possibilities of feeding with dry feed are running out and on the verge of economic profitability.

According to recent reports, liquid feed is currently being applied in many parts of the world, especially in western Europe. Approximate figures indicate that more than 60% of fatteners in Denmark and Sweden, as well as the majority of sows there, are fed liquid feed. Another example from the Netherlands and France indicated that about a third part of all pigs are fed with liquid feed, of which 50 to 60% of fatteners in the Netherlands and over 70% of fatteners in France. Around 40% of fattening pigs in Germany and far fewer sows, receive liquid feed. It is estimated that approximately 70% of pigs are fed according to this system in Ireland (Byrgesen et al. 2021).

Liquid feeding, due to the availability and low prices of maize, is less popular in North America than in Europe. The exception is Ontario, Canada, where in

2012 approximately 20% of fatteners were fed liquid feed.

It should be mentioned that in Poland there is no precise data on the number of pigs fed liquid feed. This may give the impression that the scale of the discussed issue is small. Currently, only about 100 liquid feed systems are installed in Poland. Considering the data available for several years on the effectiveness of feeding with liquid feed, not paying enough attention to the discussed issue should be a cause for reflexion.

Fermented feeds

Fermented pig feed is showing promise as an alternative to zinc oxide. According to reports, application of fermented feed can lead to improved growth of the small intestinal villi and hind gut mucosa, improve gut barrier function in the jejunum and colon (including the integrity of the mucosal and brush border, the presence of GALT structures and minimal inflammation indices), and shift the colon microbiota toward greater microbial diversity and robustness (Shi et al. 2016, Yuan et al. 2017, Zhu et al. 2017). Furthermore, fermented feeds show better digestibility because they are more easily digested by young animals. It is likely that the process of pre-fermentation of rapeseed meal through microbial control enhances its nutritional value. Studies have shown that microbial fermentation can improve the digestibility of feedstuffs such as rapeseed meal, as well as neutralises antinutritional components such as tannins, glucosinolates, and phytic acid (Satessa et al. 2020).

Bacteriophages

Since the beginning of the 20th century, “phage therapy” has been known to be applied in human and veterinary medicine (Sulakvelidze et al. 2001). It is used successfully in the treatment of certain bacterial infections. A bacteriophage is a type of virus that attacks and destroys bacterial cells by an induction process (Howard-Varona et al. 2017). It turns out that it is possible to use phages to supplement piglets. According to Zeng et al. (2021), the phage cocktail added to the food maintained the performance at the level observed in other animal groups administered antibiotics. Furthermore, supplementing an antibiotic diet with phages had no effect on serum IgA, IgG, IgM proteins used by the immune system to identify and neutralize risks (Janeway 2001), IL-2, IL-12 – signaling molecules relevant for immune defense (Gołab et al. 2007) and IFN- γ – interferon type II as macrophages activator (Schoenborn and Wilson 2007). However,

piglets fed a 400 mg/kg bacteriophage-containing diet had lower IL-1 β and TNF- α concentrations and higher serum IL-10 concentrations than antibiotic-fed piglets (Kim et al. 2014). In summary, this could mean that the bacteriophage diet had a stronger anti-inflammatory effect (Moore et al. 2001, Masters et al. 2009, Gołab et al. 2011). Earlier, it was also shown that phages can be growth promoters in pigs. Administration of 400 mg/kg bacteriophages was found to increase final body weight and reduce the incidence of diarrhea in weaning piglets compared to the antibiotic group. This means that 400 mg / kg of phage supplementation significantly improved growth performance compared to an antibiotic diet (Kim et al. 2014). There are also reports showing that phage therapy is effective against bacterial infections such as *Salmonella* spp. (Wall et al. 2009, Won et al. 2021); *E. coli* (Smith and Huggins 1983, Lee et al. 2017) and *Clostridium* spp. (Kim et al. 2017). Lee et al. (2017) in their study also argue that the use of phages could be an alternative to antibiotics and zinc compounds.

Acknowledgements

The scientific activity of prof. Grzegorz Woźniakowski regarding the epidemiology of pig infectious diseases is funded by the project of National Science Centre: UMO-2020/39/B/NZ7/00493.

References

- Abbaszadeh S, Sharifzadeh A, Shokri H, Khosravi AR, Abbaszadeh A (2014) Antifungal efficacy of thymol, arvacrol, eugenol and menthol as alternative agents to control the growth of food-relevant fungi. *J Mycol Med* 24: e51-e56.
- Ahmed ST, Hwang JA, Hoon J, Mun HS, Yang CJ (2014) Comparison of single and blend acidifiers as alternative to antibiotics on growth performance, fecal microflora, and humoral immunity in weaned piglets. *Asian-Australas J Anim Sci* 1: 93-100.
- Barba-Vidal E, Castillejos L, Roll VF, Cifuentes-Orjuela G, Moreno Muñoz JA, Martín-Orúe SM (2017) The probiotic combination of *Bifidobacterium longum* subsp. infantis CECT 7210 and *Bifidobacterium animalis* subsp. lactis BPL6 reduces pathogen loads and improves gut health of weaned piglets orally challenged with *Salmonella typhimurium*. *Front Microbiol* 8: 1570.
- Barba-Vidal E, Martin-Orue SM, Castillejos L (2019) Practical aspects of the use of probiotics in pig production: A review. *Livest Sci* 223: 84-96.
- Baschieri A, Ajvazi MD, Tonfack JL, Valgimigli L, Amorati R (2017) Explaining the antioxidant activity of some common non-phenolic components of essential oils. *Food Chem* 232: 656-663.
- Bednorz C, Oelgeschläger K, Kinnemann B, Hartmann S, Neumann K, Pieper R, Bethe A, Semmler T, Tedin K, Schierack P, Wieler LH, Guenther S (2013) The broader context of antibiotic resistance: zinc feed supplementation of piglets increases the proportion of multi-resistant *Escherichia coli* in vivo. *Int J Med Microbiol* 303: 396-403.
- Bonetti A, Tugnoli B, Piva A, Grilli E (2021) Towards zero zinc oxide: Feeding strategies to manage post-weaning diarrhea in piglets. *Animals (Basel)* 11: 642.
- Bouchra C, Achouri M, Idrissi Hassani LM, Hmamouchi M (2003) Chemical composition and antifungal activity of essential oils of seven Moroccan labiatae against *Botrytis cinerea* Pers: Fr. *Jof Ethnopharmacol* 89: 165-169.
- Brenes A, Roura E (2010) Essential oils in poultry nutrition: Main effects and modes of action. *Anim Feed Sci Technol* 158: 1-14.
- Broadley MR, White PJ, Hammond JP, Zelko I, Lux A (2007) Zinc in plants. *New Phytol* 173: 677-702.
- Burt SA, Reinders RD (2003) Antibacterial activity of selected plant essential oils against *Escherichia coli* O157:H7. *Lett Appl Microbiol* 36: 162-167.
- Byrgesen N, Madsen JG, Larsen C, Kjeldsen NJ, Cilieborg MS, Amdi C (2021) The effect of feeding liquid or dry creep feed on growth performance, feed disappearance, enzyme activity and number of eaters in suckling piglets. *Animals (Basel)* 11: 3144.
- Bøe K (1991) The process of weaning in pigs: When the sow decides. *Appl Anim Behav Sci* 30: 47-59.
- Canibe N, Højberg O, Kongsted H, Vodolazska D, Lauridsen C, Nielsen TS, Schönherz AS (2022) Review on Preventive Measures to Reduce Post-Weaning Diarrhoea in Piglets. *Animals (Basel)* 12: 2585.
- Case CL, Carlson MS (2002) Effect of feeding organic and inorganic sources of additional zinc on growth performance and zinc balance in nursery pigs. *J Anim Sci* 80: 1917-1924.
- Casewell M, Friis C, Marco E, McMullin P, Phillips I (2003) The European ban on growth-promoting antibiotics and emerging consequences for human and animal health. *J Antimicrob Chemother* 52: 159-161.
- Catalkaya G, Venema K, Lucini L, Rocchetti G, Delmas D, Daglia M, De Filippis A De, Xiao H, Quiles JL, Xiao J, Capanoglu E (2020) Interaction of dietary polyphenols and gut microbiota: Microbial metabolism of polyphenols, influence on the gut microbiota, and implications on host health. *Food Front* 1: 109-133.
- Chorianopoulos NG, Giaouris ED, Skandamis PN, Haroutounian SA, Nychas GJ (2008) Disinfectant test against monoculture and mixed-culture biofilms composed of technological, spoilage and pathogenic bacteria: bactericidal effect of essential oil and hydrosol of *Satureja thymbra* and comparison with standard acid-base sanitizers. *J Appl Microbiol* 104: 1586-1596.
- Christaki E, Bonos E, Giannenas I, Florou-Paneri PF (2021) Aromatic plants as a source of bioactive compounds. *Agriculture* 2: 228-243.
- De Martino L, De Feo V, Nazzaro F (2009) Chemical composition and in vitro antimicrobial and mutagenic activities of seven Lamiaceae essential oils. *Molecules* 14: 4213-4230.
- Del Rio D, Rodriguez-Mateos A, Spencer JP, Tognolini M, Borges G, Crozier A (2013) Dietary (poly)phenolics

- in human health: structures, bioavailability, and evidence of protective effects against chronic diseases. *Antioxid Redox Signal* 18: 1818-1892.
- Dianawati D, Mishra V, Shah NP (2016) Survival of microencapsulated probiotic bacteria after processing and during storage: A Review. *Critic Rev Food Sci Nutr* 56: 1685-1716.
- Dourmad JY, Étienne M, Valancogne A, Dubois S, Van Milgen J, Noblet J (2008) InraPorc: a model and decision support tool for the nutrition of sows. *Anim Feed Sci Technol* 143: 372-386.
- Dubreuil JD (2017) Enterotoxigenic *Escherichia coli* and probiotics in swine: what the bleep do we know? *Biosci Microbiota Food Health* 36: 75-90.
- Dumitru M, Habeanu M, Lefter NA, Gheorghe A (2020) The effect of *Bacillus licheniformis* as direct-fed microbial product on growth performance, gastrointestinal disorders and microflora population in weaning piglets. *Rom Biotechnol Lett* 25: 2060-2069.
- Ercan MT, Bor NM (1991) Phagocytosis by macrophages in zinc-deficient rats. *Int J Appl Instrum* 18: 765-768.
- European Commission. Commission Implementing Decision of 26.6.2017 Concerning, in the Framework of Article 35 of Directive 2001/82/EC of the European Parliament and of the Council, the Marketing Authorisations for Veterinary Medicinal Products Containing “Zinc Oxide” to be Ad; Official Journal of the European Union: Brussels, Belgium, 2017. www.ema.europa.eu/en/documents/report/sales-veterinary-antimicrobial-agents-31-european-countries-2019-2020-trends-2010-2020-eleventh_en.pdf
- European Commission. 2003. “Opinion of the Scientific Committee on Animal Nutrition on the Use of Zinc in Feed.” https://ec.europa.eu/food/sites/food/files/safety/docs/animal-feed_additives_rules_scan-old_report_out120.pdf
- Gammoh, NZ, Rink L (2019) Zinc and the immune system. In: *Nutrition and immunity*. Springer International Publishing, Cham Switzerland, pp 127-158.
- Goodner KL, Mahattanatawee K, Plotto A, Sotomayor J, Jordán MJ (2006) Aromatic profiles of *Thymus hyemalis* and Spanish *T. vulgaris* essential oils by GC-MS/GC-O. *Industrial Crops and Prod* 24: 264-268.
- Gołąb J, Jakóbsiak M, Lasek W, Stokłosa T. *Immunology*. Warszawa: Wydawnictwo Naukowe PWN, 2007, chapter 7 – Lymphocyte maturation pp. 108-111. ISBN 978-83-01-15154-6.
- Gołąb J, Jakóbsiak M, Lasek W, Stokłosa T. *Immunology*. Warsaw: Wydawnictwo Naukowe PWN, 2011, chapter 11 – Cytokines p. 164. ISBN 978-83-01-15154-6.
- Grilli E, Tugnoli B, Vitari F, Domeneghini C, Morlacchini M, Piva A, Prandini A (2015) Low doses of microencapsulated zinc oxide improve performance and modulate the ileum architecture, inflammatory cytokines and tight junctions expression of weaned pigs. *Animal* 9: 1760-1768.
- Halas D, Heo JM, Hansen CF, Kim JC, Hampson DJ, Mullan BP, Pluske JR (2007) Organic acids, prebiotics and protein level as dietary tools to control the weaning transition and reduce post-weaning diarrhoea in piglets. *CAB Reviews: Perspectives in Agriculture, Veterinary Science Nutrition and Natural Resources* 2: p 13.
- Hedemann MS, Jensen BB, Poulsen, HD (2006) Influence of dietary zinc and copper on digestive enzyme activity and intestinal morphology in weaned pigs. *Anim Sci* 84: 3310-3320.
- Heo JM, Kim JC, Hansen CF, Mullan BP, Hampson DJ, Maribo H, Kjeldsen N, Pluske JR (2010) Effects of dietary protein level and zinc oxide supplementation on the incidence of post-weaning diarrhoea in weaner pigs challenged with an enterotoxigenic strain of *Escherichia coli*. *Livestock Sci* 133: 210-213.
- Honscheid A, Rink L, Haase H (2009) T-Lymphocytes: A target for stimulatory and inhibitory effects of zinc ions. *Endocr Metab Immune Disord Drug Targets* 9: 132-144.
- Howard-Varona C, Hargreaves KR, Abedon ST, Sullivan MB (2017) Lysogeny in nature: mechanisms, impact and ecology of temperate phages. *International Society for Microbial Ecology* 11: 1511-1520. doi:10.1038/ismej.2017.16.
- Hrala M, Bosák J, Mícenková L, Křenová J, Lexa M, Pirková V, Tomáščíková Z, Koláčková I, Šmajš D (2021) *Escherichia coli* strains producing selected bacteriocins inhibit porcine enterotoxigenic *Escherichia coli* (ETEC) under both in vitro and in vivo conditions. *Appl Environ Microbiol* 87: e0312120
- Hu C, Song J, Li Y, Luan Z, Zhu K (2013) Diosmectite-zinc oxide composite improves intestinal barrier function, modulates expression of pro-inflammatory cytokines and tight junction protein in early weaned pigs. *Br J Nutr* 110: 681-688.
- Hu CH, Xiao K, Song J, Luan ZS (2013) Effects of zinc oxide supported on zeolite on growth performance, intestinal microflora and permeability, and cytokines expression of weaned pigs. *Anim Feed Sci Technol* 181: 65-71.
- Hu Y, Dun Y, Li S, Zhao S, Peng N, Liang Y (2014) Effects of *Bacillus subtilis* KN-42 on Growth Performance, Diarrhea and Faecal Bacterial Flora of Weaned Piglets. *Asian-Australas J Anim Sci* 27: 1131-1140.
- Hu Y, Dun Y, Li S, Zhang D, Peng N, Zhao S, Liang Y (2015) Dietary *Enterococcus faecalis* LAB31 improves growth performance, reduces diarrhea, and increases fecal *Lactobacillus* number of weaned piglets. *PLoS One* 10: e0116635.
- Højberg O, Canibe N, Poulsen HD, Hedemann MS, Jensen BB (2005) Influence of dietary zinc oxide and copper sulfate on the gastrointestinal ecosystem in newly weaned piglets. *Appl Environ Microbiol* 71: 2267-2277.
- Janeway C (2001) *Immunobiology* (5th ed.). Garland Publishing ISBN 978-0-8153-3642-6.
- Jensen P, Recén B (1989) When to wean - observations from free-ranging domestic pigs. *Appl Anim Behav Sci* 23: 49-60.
- Kim JS, Hosseindoust A, Lee SH, Choi YH, Kim MJ, Lee JH, Kwon IK, Chae BJ (2017) Bacteriophage cocktail and multi-strain probiotics in the feed for weanling pigs: Effects on intestine morphology and targeted intestinal coliforms and *Clostridium*. *Animal*, 11: 45–53.
- Kim KH, Ingale SL, Kim JS, Lee SH, Lee JH, Kwon I, Chae BJ (2014) Bacteriophage and probiotics both enhance the performance of growing pigs but bacteriophage are more effective. *Anim Feed Sci Technol* 196: 88-95.
- Kloubert V, Blaabjerg K, Dalgaard TS, Poulsen HD, Rink L, Wessels I (2018) Influence of zinc supplementation

- on immune parameters in weaned pigs. *J Trace Elem Med Biol* 49: 231-240.
- Koo B, Choi J, Yang C, Nyachoti CM (2020) Diet complexity and L-threonine supplementation: effects on growth performance, immune response, intestinal barrier function, and microbial metabolites in nursery pigs. *J Anim Sci* 98: skaa125.
- Kroismayr A, Steiner T, Zhang C (2006) Influence of a phyto-genic feed additive on performance of weaner piglets. *Anim Sci. Pub. American Society of Animal Science*, 1111 North Dunlap Ave, Savoy, IL 61874 USA, pp 270-270.
- Lan R, Koo J, Kim I (2017) Effects of *Lactobacillus acidophilus* supplementation on growth performance, nutrient digestibility, fecal microbial and noxious gas emission in weaning pigs. *J Sci Food Agric* 97: 1310-1315.
- Le Floc'h N, Seve B (2007) Biological roles of tryptophan and its metabolism: Potential implications for pig feeding. *Livest Sci*, 112: 23-32.
- Lee CY, Kim SJ, Park BC, Han JH (2017) Effects of dietary supplementation of bacteriophages against enterotoxi-genic *Escherichia coli* (ETEC) K88 on clinical symptoms of post-weaning pigs challenged with the ETEC patho-gen. *Jof Anim Physiol Anim Nutr* 101: 88-95.
- Lee SR (2018) Critical role of zinc as either an antioxidant or a prooxidant in cellular systems. *Oxid Med Cell Longev* 91: 9156285.
- Li P, Piao X, Ru Y, Han X, Xue L, Zhang H (2012) Effects of adding essential oil to the diet of weaned pigs on perfor-mance, nutrient utilization, immune response and intesti-nal health. *Asian-Australas J Anim Sci* 25: 1617-1626.
- Liao SF (2021) Invited review: Maintain or improve piglet gut health around weaning: The fundamental effects of dietary amino acids. *Animals* 11: 1110.
- Lien KA, Sauer WC, Fenton M (1997) Mucin output in ileal digesta of pigs fed a protein-free diet. *Z Ernahrungswiss* 36: 182-190.
- Liu Y, Yang X, Xin H, Chen S, Yang C, Duan Y, Yang X (2017) Effects of a protected inclusion of organic acids and essential oils as antibiotic growth promoter alterna-tive on growth performance, intestinal morphology and gut microflora in broilers. *Anim Sci J* 88: 1414-1424.
- López-Gálvez G, López-Alonso M, Pechova A, Mayo B, Dierick N, Gropp J (2021) Alternatives to antibiotics and trace elements (copper and zinc) to improve gut health and zootechnical parameters in piglets: A review. *Anim Feed Sci Technol* 271: 114727.
- Luna A, Lábague MC, Zygadlo JA, Marin RH (2010) Effects of thymol and carvacrol feed supplementation on lipid oxidation in broiler meat. *Poult Sci* 89: 366-370.
- MacDonald RS (2000) The role of zinc in growth and cell proliferation. *J Nutr* 130: 1500S-1508S.
- Mahfuz S, Shang Q, Piao X (2021) Phenolic compounds as natural feed additives in poultry and swine diets: a review. *J Anim Sci Biotechnol*, 12: 48.
- Masters SL, Simon A, Aksentijevich I, Kastner DL (2009) *Horror autoinflamaticus*: The molecular pathophysio-logy of autoinflammatory disease. *Annu Rev Immunol* 27: 621-668.
- McGilvray WD, Wooten H, Rakhshandeh AR, Petry A, Rakhshandeh A (2019) Immune system stimulation increases dietary threonine requirements for protein depo-sition in growing pigs. *J Anim Sci* 97: 735-744.
- Moore KW, de Waal Malefyt R, Coffman RL, O'Garra A (2001) Interleukin-10 and the interleukin-10 receptor. *Annu Rev Immunol* 19: 683-765.
- Moxley RA (2000) Edema disease. *Vet Clin North Am. Food Anim Pract* 16: 175-185.
- National Research Council (2012) *Nutrient Requirements of Swine: Eleventh Revised Edition*. Washington, DC: The National Academies Press, p 420.
- Nazzaro F, Fratianni F, de Martino L, Coppola R, de Feo V (2013) Effect of essential oils on pathogenic bacteria. *Pharmaceuticals* 12: 1451-1474.
- Nyachoti CM, Omogbenigun FO, Rademacher M, Blank G (2006) Performance responses and indicators of gastroin-estinal health in early-weaned pigs fed low-protein amino acid-supplemented diets. *J Anim Sci* 84:125-134.
- Ociepa-Kubicka A, Ociepa E (2012) *Toxic impact of heavy metals on plants, animals and humans*. Published by the the Czestochowa University of Technology. Engineering and Environmental Protection. vol. 15, no. 2. 169-180.
- Park SY, Bok SH, Jeon SM, Park YB, Lee SJ, Jeong TS, Choi MS (2002) Effect of rutin and tannic acid supple-ments on cholesterol metabolism in rats. *Nutri Res* 22: 283-295.
- Pasquet J, Chevalier Y, Pelletier J, Couval E, Bouvier D, Bolzinger MA (2014) The contribution of zinc ions to the antimicrobial activity of zinc oxide. *Colloids and Surfaces A: Physicochemical and Engineering Aspects* 457: 263-274.
- Pearce SC, Sanz Fernandez MV, Torrison J, Wilson ME, Baumgard LH, Gabler NK (2015) Dietary organic zinc attenuates heat stress-induced changes in pig intestinal integrity and metabolism. *Anim Sci* 93: 4702-4713.
- Pieper R, Villodre Tudela C, Taciak M, Bindelle J, Pérez JF, Zentek J (2016) Health relevance of intestinal protein fermentation in young pigs. *Anim Health Res Rev* 17: 137-147.
- Poulsen HD (1998) Zinc and copper as feed additives, growth factors or unwanted environmental factors. *J Anim Feed Sci* 7 (Suppl 1): 135-142.
- Prakash A, Bharti K, Majeed AB (2015) Zinc: Indications in brain disorders. *Fundam Clin Pharmacol* 29: 131-149.
- Prasad AS (2008) Zinc in human health: effect of zinc on im-mune cells. *Mol Med* 14:353-357.
- Puvača N, Stanačev V, Glamočić D, Lević J, Perić L, Stanače, V, Milić D (2013) Beneficial effects of phyto-additives in broiler nutrition. *World's Poult Sci J* 69: 27-34.
- Quideau S, Deffieux D, Douat-Casassus C, Pouységu L (2011) Plant polyphenols: chemical properties, biological activi-ties, and synthesis. *Angew Chem Int Ed* 50: 586-621.
- Ren M, Liu XT, Wang X, Zhang GJ, Qiao SY, Zeng XF (2014) Increased levels of standardized ileal digestible threonine attenuate intestinal damage and immune responses in *Escherichia coli* K88+ challenged weaned piglets. *Anim Feed Sci Technol* 195: 67-75.
- Rhouma M, Fairbrother JM, Beaudry F, Letellier A (2017) Post weaning diarrhea in pigs: risk factors and non-colis-tin-based control strategies. *Acta Vet Scand* 59: 31.
- Rolles B, Maywald M, Rink L (2018) Influence of zinc defi-ciency and supplementation on NK cell cytotoxicity. *J Funct Foods* 48: 322-328.
- Roselli M, Finamore A, Garaguso I, Britti MS, Mengheri E

- (2003) Zinc Oxide Protects Cultured Enterocytes from the Damage Induced by *Escherichia coli*. *J Nutr* 133: 4077-4082.
- Sargeant HR, Miller HM, Shaw MA (2011) Inflammatory response of porcine epithelial IPEC J2 cells to enterotoxigenic *E. coli* infection is modulated by zinc supplementation. *Mol Immunol* 48: 2113-2121.
- Satessa GD, Tamez-Hidalgo P, Hui Y, Cieplak T, Krych L, Kjærulff S, Brunsgaard G, Nielsen DS, Nielsen MO (2020) Impact of Dietary Supplementation of Lactic Acid Bacteria Fermented Rapeseed with or without Macroalgae on Performance and Health of Piglets Following Omission of Medicinal Zinc from Weaner Diets. *Animals (Basel)* 10: 137.
- Schoenborn JR, Wilson CB (2007) Regulation of interferon-gamma during innate and adaptive immune responses. *Adv Immunol* 96 :41-101.
- Shi C, He J, Wang J, Yu J, Yu B, Mao X, Zheng P, Huang Z, Chen D (2016) Effects of *Aspergillus niger* fermented rapeseed meal on nutrient digestibility, growth performance and serum parameters in growing pigs. *Anim Sci J* 87: 557-563.
- Shu Q, Qu F, Gill HS (2001) Probiotic treatment using *Bifidobacterium lactis* HN019 reduces weanling diarrhea associated with rotavirus and *Escherichia coli* infection in a piglet model. *J Pediatr Gastroenterol Nutri* 33: 171-177.
- Si W, Gong J, Tsao R, Zhou T, Yu H, Poppe C, Johnson R, Du Z (2006) Antimicrobial activity of essential oils and structurally related synthetic food additives towards selected pathogenic and beneficial gut bacteria. *J Appl Microbiol* 100: 296-305.
- Slifierz MJ, Friendship R, Weese JS (2015) Zinc oxide therapy increases prevalence and persistence of methicillin-resistant staphylococcus aureus in pigs: A randomized controlled trial. *Zoonoses Public Health* 62: 301-308.
- Smith HW, Huggins MB (1983) Effectiveness of phages in treating experimental *Escherichia coli* diarrhoea in calves, piglets and lambs. *J Gen Microbiol* 129: 2659-2675.
- Söderberg, TA, Sunzel B, Holm S, Elmros T, Hallmans G, Sjöberg S (1990) Antibacterial effect of zinc oxide in vitro. *Scand J Plast Surg Hand Surg* 24: 193-197.
- Suiryanrayna MV, Ramana JV (2015) A review of the effects of dietary organic acids fed to swine. *J Anim Sci and Biotechnol* 6: 45.
- Sulakvelidze A, Alavidze Z, Morris JR (2001) Bacteriophage therapy. *Antimicrob Agents Chemothe* 45: 649-659.
- Suo C, Yin Y, Wang X, Lou X, Song D, Wang X, Gu Q (2012) Effects of *Lactobacillus plantarum* ZJ316 on pig growth and pork quality. *BMC Vet Res* 8: 89.
- Sutaphanit P, Chitprasert P (2014) Optimisation of microencapsulation of holy basil essential oil in gelatin by response surface methodology. *Food Chem* 150: 313-320.
- Tan C, Wei H, Sun H, Ao J, Long G, Jiang S, Peng J (2015) Effects of dietary supplementation of oregano essential oil to sows on oxidative stress status, lactation feed intake of sows, and piglet performance. *BioMed Res Int* 2015: 525218.
- Tian QY, Piao XS (2019) Essential Oil Blend Could Decrease Diarrhea Prevalence by Improving Antioxidative Capability for Weaned Pigs. *Animals (Basel)*. 9: 847.
- Trevisi P, Corrent E, Mazzoni M, Messori S, Priori D, Gherpelli Y, Simongiovanni A, Bosi P (2015) Effect of added dietary threonine on growth performance, health, immunity and gastrointestinal function of weaning pigs with differing genetic susceptibility to *Escherichia coli* infection and challenged with *E. coli* K88ac. *J Anim Physiol Anim Nutr* 99: 511-520.
- Tsiloyiannis VK, Kyriakis SC, Vlemmas J, Sarris K (2001) The effect of organic acids on the control of post-weaning oedema disease of piglets. *Res Vet Sci* 70: 281-285.
- Vahjen W, Pietruszyńska D, Starke IC, Zentek J (2015) High dietary zinc supplementation increases the occurrence of tetracycline and sulfonamide resistance genes in the intestine of weaned pigs. *Gut Pathog* 7: 23.
- Van Milgen J, Dourmad JY (2015) Concept and application of ideal protein for pigs. *J Anim Sci Biotechnol* 6: 15.
- Van Milgen J, Valancogne A, Dubois S, Dourmad JY, Sève B, Noblet J (2008) InraPorc: a model and decision support tool for the nutrition of growing pigs. *Anim Feed Sci Technol* 143: 387-405.
- Wall SK, Zhang J, Rostagno MH, Ebner PD (2010) Phage therapy to reduce preprocessing *Salmonella* infections in market-weight swine. *Appl Environ Microbiol* 76: 48-53.
- Wan K, Li Y, Sun W, An R, Tang Z, Wu L, Chen H, Sun Z (2020) Effects of dietary calcium pyruvate on gastrointestinal tract development, intestinal health and growth performance of newly weaned piglets fed low-protein diets. *J Appl Microbiol* 128: 355-365.
- Wang T, Teng K, Liu Y, Shi W, Zhang J, Dong E, Zhang X, Tao Y, Zhong J (2019) *Lactobacillus plantarum* PFM 105 Promotes Intestinal Development Through Modulation of Gut Microbiota in Weaning Piglets. *Front Microbiol* 10: 90.
- Wang WW, Qiao SY, Li DF (2009) Amino acids and gut function. *Amino Acids*. 37: 105-110.
- Wang Y, Wu Y, Wang B, Cao X, Fu A, Li Y, Li W (2017) Effects of probiotic *Bacillus* as a substitute for antibiotics on antioxidant capacity and intestinal autophagy of piglets. *AMB Express* 7: 52.
- Wei HK, Xue HX, Zhou ZX, Peng J (2017) A carvacrol-thymol blend decreased intestinal oxidative stress and influenced selected microbes without changing the messenger RNA levels of tight junction proteins in jejunal mucosa of weaning piglets. *Animal* 2: 193-201.
- Wellock IJ, Fortomaris PD, Houdijk JG, Kyriazakis I (2008) Effects of dietary protein supply, weaning age and experimental enterotoxigenic *Escherichia coli* infection on newly weaned pigs. *Health Animal* 2: 834-842.
- Won YK, Kim SJ, Han JH (2021) The protective effect of dietary supplementation of *Salmonella*-specific bacteriophages in post-weaning piglets challenged with *Salmonella typhimurium*. *J Adv Vet Anim Res* 8: 440-447.
- Woźniakowska A, Kozera W, Karpiesiuk K (2017) Organic acids in piglet nutrition. *Med Weter* 73: 76-81.
- Yang Z, Wang Y, He T, Bumbie G, Wu L, Sun Z, Sun W, Tang Z (2021) Effects of dietary *Yucca Schidigera* Extract and oral *Candida utilis* on growth performance and intestinal health of weaned piglets. *Front Nutr* 8: 685540.
- Yin J, Li X, Li D, Yue T, Fang Q, Ni J, Zhou X, Wu G (2009) Dietary supplementation with zinc oxide stimulates ghrelin secretion from the stomach of young pigs. *J Nutr Biochem* 20: 783-790.

- Yuan L, Chang J, Yin Q, Lu M, Di Y, Wang P, Wang Z, Wang E, Lu F (2017) Fermented soybean meal improves the growth performance, nutrient digestibility, and microbial flora in piglets. *Anim Nutr* 3: 19–24.
- Yue LY, Qiao SY (2008) Effects of low-protein diets supplemented with crystalline amino acids on performance and intestinal development in piglets over the first 2 weeks after weaning. *Livest Sci* 115: 144-152.
- Zeghib A, Kabouche A, Laggoune S, et al (2017) Antibacterial, antiviral, antioxidant and antiproliferative activities of *Thymus guyonii* essential Oil. *Natural Product Communications* 12: 10.
- Zeng Y, Wang Z, Zou T, Chen J, Li G, Zheng L, Li S, You J (2021) Bacteriophage as an alternative to antibiotics promotes growth performance by regulating intestinal inflammation, intestinal barrier function and gut microbiota in weaned piglets. *Front Vet Sci* 8: 623899.
- Zeng Z, Xu X, Zhang Q, Li P, Zhao P, Li Q, Liu J, Piao X (2015) Effects of essential oil supplementation of a low-energy diet on performance, intestinal morphology and microflora, immune properties and antioxidant activities in weaned pigs. *Anim Sci J* 86: 279-285.
- Zeyner A, Boldt E (2006) Effects of a probiotic *Enterococcus faecium* strain supplemented from birth to weaning on diarrhoea patterns and performance of piglets. *J Anim Physiol Anim Nutri* 90: 25-31.
- Zhang B, Guo Y (2009) Supplemental zinc reduced intestinal permeability by enhancing occludes and zonula occludes protein-1 (ZO-1) expression in weaning piglets. *Br J Nutri* 102: 687-693.
- Zhang H, Chen Y, Li Y, Zhang T, Ying Z, Su W, Zhang L, Wang T (2019) l-Threonine improves intestinal mucin synthesis and immune function of intrauterine growth-retarded weanling piglets. *Nutrition* 59: 182-187.
- Zhang W, Bao C, Wang J, Zang J, Cao Y (2020) Administration of *Saccharomyces boulardii* mafic-1701 improves feed conversion ratio, promotes antioxidant capacity, alleviates intestinal inflammation and modulates gut microbiota in weaned piglets. *J Anim Sci Biotechnol* 11: 112.
- Zhang W, Zhu YH, Zhou D, Wu Q, Song D, Dicksved J, Wang JF (2017) Oral Administration of a select mixture of *Bacillus* probiotics affects the gut microbiota and goblet cell function following *Escherichia coli* challenge in newly weaned pigs of genotype MUC4 that are supposed to be enterotoxigenic *E. coli* F4ab/ac receptor negative. *Appl Environ Microbiol* 83: e02747-16.
- Zhang Y, Wu T, Chen Z, Meng Y, Zhu Z, Wang Q, Tian J, Yi D, Wang L, Zhao D, Hou Y (2021) Dietary supplementation with *enterococcus faecium* R1 attenuates intestinal and liver injury in piglets challenged by lipopolysaccharide. *Animals (Basel)* 11: 1424.
- Zhaxi Y, Meng X, Wang W, Duan-Nai-An A (2020) Yeast probiotic, improves intestinal mucosa integrity and immune function in weaned piglets. *Sci Rep* 10: 4556.
- Zhu C, Lv H, Chen Z, Wang L, Wu X, Chen Z, Zhang W, Liang R, Jiang Z (2017) Dietary zinc oxide modulates antioxidant capacity, small intestine development, and jejunal gene expression in weaned piglets. *Biol Trace Elem Res* 175: 331-338.
- Zhu J, Gao M, Zhang R, Sun Z, Wang C, Yang F, Huang T, Qu S, Zhao L, Li Y, Hao Z (2017) Effects of soybean meal fermented by *L. plantarum*, *B. subtilis* and *S. cerevisiae* on growth, immune function and intestinal morphology in weaned piglets. *Microb Cell Fact* 16: 191.
- Zou Y, Wang J, Peng J, Wei H (2016) Oregano essential oil induces SOD1 and GSH expression through Nrf2 activation and alleviates hydrogen peroxide-induced oxidative damage in IPEC-J2 cells. *Oxid Med and Cell Longev* 2016: 5987183.