Polish Journal of Veterinary Sciences Vol. 18, No. 4 (2015), 881–883



DOI 10.1515/pjvs-2015-0115

Short communication

# A serological and molecular study on the occurrence of mycoplasmas in European bison (*Bison bonasus*) from two areas of Eastern Poland

K. Dudek<sup>1</sup>, D. Bednarek<sup>1</sup>, E. Szacawa<sup>1</sup>, R.D. Ayling<sup>2</sup>, M.K. Krzysiak<sup>3</sup>, J. Marczuk<sup>4</sup>

## **Abstract**

European bison (*Bison bonasus*) from two different areas of Eastern Poland showing gross pathology possibly associated with mycoplasma infections were tested for ruminant *Mycoplasma* species using serological and molecular methods. Fifty-five samples, blood or tissue were collected from 28 animals during 2013-2014. Six sera were positive for *Mycoplasma bovis*. The ELISA and complement fixation test for *Mycoplasma mycoides* subsp. *mycoides* gave a few weak reactions, but were negative by immunoblotting and molecular methods.

**Key words:** European bison, ruminant mycoplasma, serology, molecular biology

# Introduction

Approximately 1432 European bison (*Bison bonasus*) are present in Poland. Nearly 60% of them populate two areas of Eastern Poland: the Polish region of the Białowieża Forest (BF) with 559 heads; and the Bieszczady Mountains (BM) with 301 heads (Raczyński 2015). *M. bovis* has previously been linked with disease in bison. In Poland serological evidence in one bison (Krzysiak et al. 2014); pneumonia, laryn-

gitis, arthritis, synovitis and tenosynovitis in feedlot bison (Dyer et al. 2008); arthritis, pleuritis and abscesses in the lung with mortality above 27% (Janardhan et al. 2010); fatal reproductive disorders and abortion in American bison in which bronchopneumonia, pleuritis and coagulation necrosis in the lung were also observed (Register et al. 2013b). Other ruminant *Mycoplasma* species that may cause respiratory disorders and arthritis in domestic and wild ruminants include: *Mycoplasma mycoides* subsp.

882 K. Dudek et al.

mycoides (Mmm), the causative agent of contagious bovine pleuropneumonia (CBPP), Mycoplasma agalactiae, and Mycoplasma capricolum subsp. capripneumoniae (Mccp) (Nicholas et al. 2008). Specific antibodies to these mycoplasmas in bison have not been observed (Krzysiak et al. 2014). As gross pathology in European bison has raised the suspicion of M. bovis infection, we assessed the current situation and extended the testing to include molecular analyses that would identify the most important ruminant mycoplasma species.

#### **Materials and Methods**

Twenty eight European bison originating from BF and BM were examined in this study. Routine necropsy as decided by the Polish Authority was carried out on three animals which had died, three which were immobilized, and 21 which were sacrificed. 55 samples were examined which comprised of 27 sera, 20 lungs, 3 livers, 1 kidney and 4 nasal swabs. The sera were tested for antibodies to M. bovis, M. agalactiae, Mmm and Mccp using the M. bovis ELISA kit (Bio-X Diagnostics, Belgium), M. agalactiae Antibody Test Kit (IDEXX), latex agglutination tests for Mccp (CapriLAT, APHA, UK) and for Mmm (BoviLAT, APHA, UK). A Mmm Antibody Test Kit (IDEXX); complement fixation test (CFT) for CBPP (Cirad, France); and immunoblotting method as described by the OIE (2012) were also used for Mmm. Mycoplasmal DNA was extracted from organ and nasal swab samples using the QIAmp DNA Mini Kit (Oiagen, Germany). The polymerase chain reaction/denaturing gradient gel electrophoresis (PCR/DGGE) method of McAuliffe et al. (2005) with modifications was used and gels analysed in comparison with the following mycoplasma controls: M. bovis, Mmm, M. bovigenitalium, M. canis, M. dispar, M. arginini, M. canadense, M. alkalescens, M. agalactiae, Mccp, and M. mycoides subsp. capri. For M. bovis DNA and antigen species-specific PCR (Subramaniam et al. 1998) and Pulmotest Mycoplasma bovis ELISA kit (Bio-X Diagnostics, Belgium) were used respectively.

# **Results and Discussion**

Distinct respiratory lesions including pneumonia, emphysema, pleural adhesions, bronchus oedema with fibrinous exudation and caseous foci were observed in 21 European bison. An ovary cyst and nodular focus in the joint was also observed in one bison. None of the respiratory lesions were typical for CBPP, but indicative of *M. bovis* infection. From 27 sera, six were positive for *M. bovis*. The *Mmm* ELISA gave

four positive and two doubtful results, one was also doubtful in the CFT. However, none were confirmed by immunoblotting or BoviLAT; and no Mmm was detected using molecular tests. These false serological reactions are likely to be due to M. bovis cross-reacting antibodies (Goncalves et al. 2008), therefore these bison do not have CBPP. No antibodies to Mccp and M. agalactiae were detected, which corresponds with our previous work (Krzysiak et al. 2014). Only six sera were positive for M. bovis. All seropositive bison originated from BF and showed pneumonia, emphysema and pleural adhesions. Unfortunately, these results were not confirmed by other methods. The Bio-X M. bovis ELISA adopted in this study is intended for use in cattle; however, Register et al. (2013a) demonstrated its application for use in bison when they compared it to their in-house ELISA designed for testing bison with agreement at more than 96%. That difference related to two samples which came from bison with an unknown history of exposure to M. bovis. In this study, a lack of detection of M. bovis antigen, despite gross pathology in seropositive bison, may indicate previous exposure to the infection, possibly from contact with infected cattle, rather than current infection. In Podlaskie province of Poland where BF is situated, M. bovis seroprevalence in cattle was assessed at 77% (Bednarek et al. 2012, Dudek and Bednarek 2012). Although the usefulness of the Bio-X M. bovis ELISA in bison testing was demonstrated earlier (Register et al. 2013a) the possibility of false positive results in this study cannot be excluded. Nevertheless it has been previously demonstrated that M. bovis does infect bison and can cause serious disease problems (Dyer et al. 2008, Janardhan et al. 2010, Register et al. 2013b). Further studies are required to show the true mycoplasma status of bison and to determine the cause of the lesions observed.

## Acknowledgements

The study was supported by Project NR12-0126-10/2011 funded by the Ministry of Science and Higher Education.

# References

Bednarek D, Ayling RD, Nicholas RA, Dudek K, Szymańska-Czerwińska M (2012) Serological survey to determine the occurrence of respiratory *Mycoplasma* infections in the Polish cattle population. Vet Rec 171: 45. Dudek K, Bednarek D (2012) Last survey of *Mycoplasma bovis* prevalence in Polish cattle affected with respiratory syndrome. Bull Vet Inst Pulawy 56: 447-451.





Dyer N, Hansen-Lardy L, Krogh D, Schaan L, Schamber E (**2008**) An outbreak of chronic pneumonia and polyarthritis syndrome caused by *Mycoplasma bovis* in feedlot bison (*Bison bison*). J Vet Diagn Invest 20: 369-371.

A serological and molecular study on the occurrence...

- Goncalves R, Regalla J, Ayling RD, Nicholas RA (2008) Impact of *Mycoplasma bovis* infection on serosurveillance for contagious bovine pleuropneumonia. Vet Rec 163: 632-633.
- Janardhan KS, Hays M, Dyer N, Oberst RD, Debey BM (2010) *Mycoplasma bovis* outbreak in a herd of North American bison (*Bison bison*). J Vet Diagn Invest 22: 797-801.
- Krzysiak MK, Dudek K, Krajewska M, Bednarek D, Szulowski K (2014) Serological studies to determine the occurrence of Johne;s disease and mycoplasma infection in the Northern-East Polish population of European bison (*Bison bonasus*). Pol J Vet Sci 17: 721-723.
- McAuliffe L, Ellis RJ, Lawes JR, Ayling RD, Nicholas RA (2005) 16S rDNA PCR and denaturing gradient gel electrophoresis; a single generic test for detecting and differentiating *Mycoplasma* species. J Med Microbiol 54: 731-739.

- Nicholas R, Ayling R, McAuliffe L (2008) Mycoplasma diseases of ruminants. 1st ed., CAB International, Wallingford.
- OIE (2012) Contagious bovine pleuropneumonia. In: Manual of diagnostic tests and vaccines for terrestrial animals. 7th ed., 2012. Chapter 2.4.9. OIE, Paris, pp 704-716.
- Raczyński J (2015) European Bison Pedigree Book 2014. 1st ed., Białowieski Park Narodowy, Białowieża.
- Register KB, Sacco RE, Olsen SC (**2013a**) Evaluation of enzyme-linked immunosorbent assays for detection of *Mycoplasma bovis*-specific antibody in bison sera. Clin Vaccine Immunol 20: 1405-1409.
- Register KB, Woodbury MR, Davies JL, Trujillo JD, Perez-Casal J, Burrage PH, Clark EG, Windeyer MC (2013b) Systemic mycoplasmosis with dystocia and abortion in a North American bison (*Bison bison*) herd. J Vet Diagn Invest 25: 541-545.
- Subramaniam S, Bergonier D, Poumarat F, Capaul S, Schlatter Y, Nicolet J, Frey J (1998) Species identification of *Mycoplasma bovis* and *Mycoplasma agalactiae* based on the *uvr*C genes by PCR. Mol Cell Probes 12: 161-169.